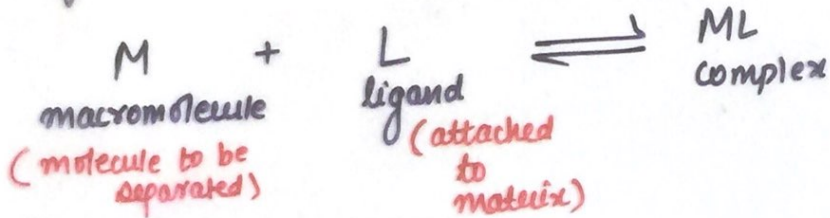


## AFFINITY CHROMATOGRAPHY.

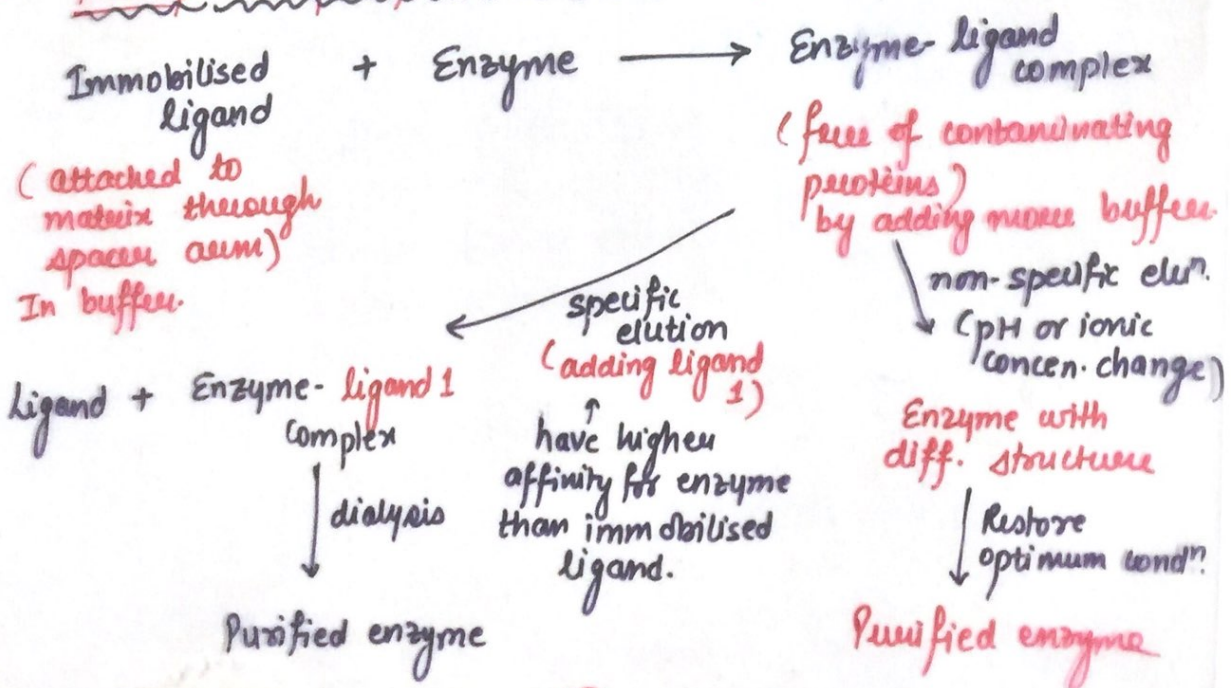
Separation is based on the specific biological interaction. Interaction is b/w a stationary immobilised ligand and a mobile liquid phase.

The technique requires that the material to be separated is capable of binding reversibly to a specific ligand that is attached to an insoluble matrix.



In the case of an enzyme, the ligand may be substrate, a competitive inhibitor or an allosteric activator. The cond<sup>n</sup>. chosen would normally be those that are optimal for enzyme-ligand binding.

### \* Principle of purification of an Enzyme.



- pH change by using dil. acetic acid & NH<sub>4</sub>OH resulting in the change in state of ionisation of groups in the ligand or the macromolecule that are critical to ligand-macromolecule binding.

### \* Characteristics of matrix for affinity chromatography

- Possess suitable and sufficient chemical groups to which the ligand may be covalently coupled.
- Interact only weakly with other macromolecules to minimise non-specific adsorption.
- Broad range of physical, chemical and thermal stability.
- Exhibit good flow properties.

Commonly used matrix :- Cross-linked dextrans and agar materials  
polyacrylamide, polymethacrylate, polystyrene, cellulose, porous glass & s

Agarose and polyacrylamide are useful matrix but they exhibit minimal adsorp<sup>n</sup>, maintain good flow prop after coupling & tolerate the extremes of pH & ionic strength as well as change in concn. of denaturants & are often needed for successful elution.

→ Polyacrylamide - is not as generally useful as agarose it fails to work properly for molecules or having a high molecular wt.

Ligand  
Ligand displays absolute specificity & group specificity.  
(it will bind exclusively to 1 particular compd.)  
(it will bind to a class of related grp. of a compd. possess a similar chemis)

②

## → Examples of group-specific ligands

<u>ligand</u>	<u>affinity</u>
5' AMP	NAD <sup>+</sup> dependent dehydrogenases, Some Kinases.
2', 5'-ADP	NADP <sup>+</sup> dependent dehydrogenases
Heparin	Lipoproteins, lipases, coagulation factors, DNA polymerases.
Proteins A and G.	Immunoglobulins.
Poly (A)	RNA containing poly (U) sequences, RNA-specific proteins.
Lysine	rRNA
Cibacron-blue	Coagulation factors, kinases, dehydrogenase, DNA polymerase, nitrate reductase.

→ It is essential that the ligand possess a suitable chemical group which can be used to attach the ligand to the matrix (not in binding of macromolecule). The most common of such groups are -NH<sub>2</sub>, -COOH, -SH and -OH.

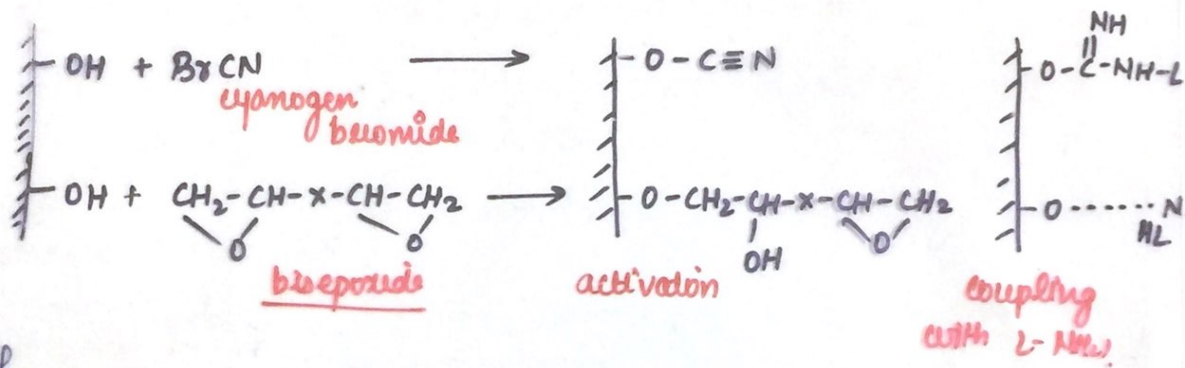
→ In case of some small ligands, steric interference occurs bcoz ligand is too near the matrix surface. This problem is usually eliminated by inserting a 6-C atom spacer b/w matrix & the ligand. This is called spacer-arm.

e.g. 6-amino hexanoic acid, 1,6 diamino hexane and (having free -COOH grp.) (having free -NH<sub>2</sub> group).

1,4 bis (2,3-epoxypropoxy) butane.

(contains free oxirane groups which allow linkage of sugars, carbohydrates or any ligand containing a -OH, -NH<sub>2</sub> or thiol group).

→ Methods of attachment of the ligand to the matrix.  
 by activating matrix done by treatment with  
CNBr, bio-epoxides, dichlorobenzotriazine, sulphonyl chloride,  
sodium periodate, N,N'-disubstituted carbodiimides.

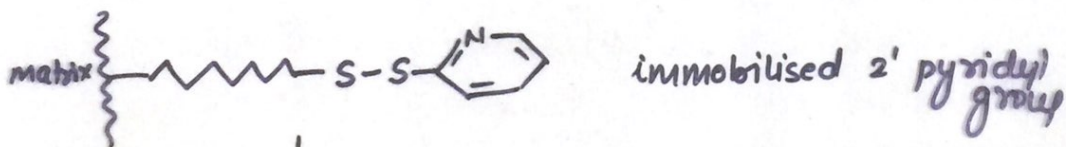


functional hydroxyl group in matrix + activating reagent → activated matrix contains reactive groups or good leaving groups. → Immobilised ligand L.

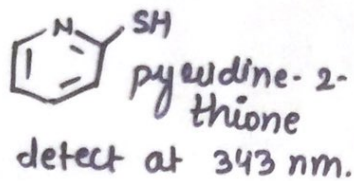
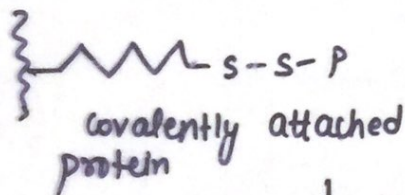
\* Covalent chromatography.

used to separate thiol-containing proteins by interaction with immobilised ligand containing a disulphide group attached to an agarose matrix.

Ligand used is disulphide 2' pyridyl group.



↓ P-SH  
thiol containing protein



- non thiol containing contaminants r eluted
- Remove unreacted thiopyridyl grp. by using 4mm dithiothreitol or mercaptoethanol.
- displace bound protein with 20-50mm DTT. (4)